

Available online at www.sciencedirect.com



Tetrahedron

Tetrahedron 64 (2008) 1494-1505

www.elsevier.com/locate/tet

## Synthesis of $(\pm)$ -9-[*c*-4, *t*-5-bis(hydroxymethyl)cyclopent-2-en-*r*-1-yl]-9*H*-adenine (BCA) derivatives branched at the 4'-position based on intramolecular S<sub>H</sub>2' cyclization

Hiroki Kumamoto<sup>a,\*</sup>, Nonoko Takahashi<sup>a</sup>, Tomomi Shimamura<sup>a</sup>, Hiromichi Tanaka<sup>a</sup>, Kazuo T. Nakamura<sup>a</sup>, Takayuki Hamasaki<sup>b</sup>, Masanori Baba<sup>b</sup>, Hiroshi Abe<sup>c</sup>, Masahiko Yano<sup>d</sup>, Nobuyuki Kato<sup>d</sup>

 <sup>a</sup> School of Pharmaceutical Sciences, Showa University, 1-5-8 Hatanodai, Shinagawa-ku, Tokyo 142-8555, Japan
<sup>b</sup> Division of Antiviral Chemotherapy, Center for Chronic Viral Diseases, Graduate School of Medical and Dental Sciences, Kagoshima University, Kagoshima 890-8544, Japan

<sup>c</sup> Nanomedical Engineering Laboratory, Discovery Research Institute, Riken, 2-1 Hirosawa, Wako, Saitama 351-0198, Japan <sup>d</sup> Department of Molecular Biology, Okayama University Graduate School of Medicine, Dentistry, and Pharmaceutical Sciences, 2-5-1 Shikata-cho, Okayama 700-8558, Japan

Received 11 October 2007; received in revised form 12 November 2007; accepted 13 November 2007 Available online 17 November 2007

#### Abstract

Synthesis of 4'-branched BCA analogues (5) was carried out. Stereospecific construction of the cis-disposed 4'-carbon-substituents and 5'-hydroxymethyl group was secured by employing the bicyclo[3.3.0]lactone **16** as a key intermediate, which was prepared by radical-mediated intramolecular  $S_H2'$  cyclization of the phenylselenomethyl ester **15**. After manipulation of the double bond of **16**, bis(Boc)adenine was introduced based on the Mitsunobu reaction of the allyl alcohol **24**. Transformation of the lactone function of **27** allowed preparation of the 4'-hydroxymethyl (**31**), the 4'-vinyl (**32**), the 4'-cyano (**34**), and the 4'-ethynyl (**35**) derivatives. Anti-HIV and anti-HCV activities of the free nucleosides **36**–**38** were also examined.

© 2007 Elsevier Ltd. All rights reserved.

Keywords: Carbocyclic nucleoside; Radical cyclization; Mitsunobu reaction

## 1. Introduction

Since the discovery of carbovir  $(1)^1$  and its prodrug abacavir  $(2)^2$  as anti-HIV agents, carbocyclic nucleosides have been recognized as a source of new antiviral agents. An anti-HIV active compound  $(\pm)$ -9-[*c*-4, *tert*-5-bis(hydroxymethyl)cyclopent-2-en-*r*-1-yl]-9*H*-adenine [ $(\pm)$ -BCA (4)] synthesized by Katagiri et al.<sup>3</sup> was designed as a hybrid of 1 and COXT (3),<sup>4</sup> which is a carbocyclic analogue of antibiotic oxetanocin.<sup>5</sup>

On the other hand, nucleosides having a 4'-carbon-substituent have attracted much attention due to the reported potent anti-HIV activity of 4'-cyanothymidine (6) and 4'-ethynyl-2'deoxycytidine (7).<sup>6</sup> Also, in our recent study, the 4'-ethynyl analogue 9 (4'-Ed4T)<sup>7</sup> of anti-HIV agent stavudine (8, d4T)<sup>8</sup> was found to show a higher activity against HIV than the parent compound 8. Based on these facts, we were interested in synthesizing 4'-branched ( $\pm$ )-BCA derivatives (5) that can be regarded as a hybrid of 4 and 4'-carbon-substituted nucleosides (6, 7, and 9).

## 2. Result and discussion

A synthetic plan for the title compounds is depicted in Scheme 1 as a retrosynthetic analysis. Manipulation of the lactone function of  $\mathbf{I}$  ensures cis-disposition between the

<sup>\*</sup> Corresponding author. Tel.: +81 3 3784 8187; fax: +81 3 3784 8252. *E-mail address:* kumamoto@pharm.showa-u.ac.jp (H. Kumamoto).



5'-hydroxymethyl group and a variety of 4'-carbon-substituents (R in 5). Compound I can be prepared by condensation of adenine base with the allyl alcohol II, which would be obtained from III by a series of reactions: epoxidation, ring opening with a selenide anion, and selenoxide elimination.



Scheme 1. Retrosynthesis of 5.

The bicyclic system of **III** could be constructed by radicalmediated 5-*exo-trig* cyclization of the phenylselenomethyl ester **IV**, which would take place through an  $S_H2'$  process with elimination of  $S(O)_n$ Ph (n=0 or 2) radical.<sup>9</sup> We anticipated that the enone **10**<sup>10</sup> can be employed for the preparation of **IV**.

#### 2.1. Radical-mediated 5-exo-trig cyclization of 14 and 15

Preparation of the substrates (14 and 15) for the radical cyclization was performed as shown in Scheme 2. The enone  $10^{10}$  was transformed to the phenylthio derivative 11 by 1,4addition of PhSH. Its conversion to the allyl sulfide **12** was performed by initial reduction of the ketone followed by trifluoromethanesulfonylation of the resulting secondary alcohol, and then by  $\beta$ -elimination of the triflate with DBN. The allyl sulfone **13** was also prepared by *m*-CPBA oxidation of **12**. After saponification of **12** and **13**, the radical precursors **14** and **15** were prepared in 68 and 81% yields, respectively, by treatment with PhSeCH<sub>2</sub>Cl<sup>11</sup> in the presence of *i*-Pr<sub>2</sub>NEt and NaI.



Scheme 2. Reagents and conditions: (a) PhSH,  $Et_3N$ ,  $CH_2Cl_2$ ; (b) NaBH<sub>4</sub>, MeOH; (c) Tf<sub>2</sub>O, pyridine,  $CH_2Cl_2$ ; (d) DBN, MeCN (87% from **10**); (e) *m*-CPBA,  $CH_2Cl_2$  (96%); (f), KOH, MeOH,  $H_2O$ ; (g) PhSeCH<sub>2</sub>Cl, NaI, *i*-Pr<sub>2</sub>NEt, DME (68% for **14**, 81% for **15**).

Radical-mediated cyclization of 14 was first carried out by adding a mixture of Bu<sub>3</sub>SnH and AIBN over 5 h to a refluxing toluene solution of 14 containing *i*-Pr<sub>2</sub>NEt (Scheme 3). Although the lactone 16 was obtained as a major product in 61% yield, this reaction also gave the saturated lactone 17 (5%) and the non-cyclized doubly reduced product 18 (18%). In contrast to this, the reaction of 15 under the same reaction conditions gave 16 in a much higher yield of 94%.



Scheme 3. Radical reactions of 14 and 15.

Although the observed different outcome between the sulfide 14 and the sulfone 15 in this cyclization reaction could be, in part, due to the lower electrophilicity of the  $\pi$  bond of the former as compared with that of the latter,<sup>12</sup> at least two factors are likely to be involved in the formation of the byproducts (17 and 18) from 14.

One is susceptibility of the phenylthio group in the allylic position to the attack of a tin radical. When **12**, lacking the phenylseleno group, was reacted under the same reaction conditions, a 15% yield of **18** was obtained, apparently via radical reduction of the phenylthio group with  $Bu_3SnH$  (Scheme 4). In contrast, the corresponding sulfone derivative **13** completely

survived in these reaction conditions. It has been reported that PhSH serves as an excellent hydrogen donor in the reaction with carbon-centered radicals, whereas benzenesulfinic acid does not.<sup>13</sup> The other factor is an efficient reducing character of the generated PhSH to radical intermediates. In fact, when cyclization reaction of the allyl sulfone **15** was carried out in the presence of PhSH (0.5 equiv), the methyl ester **13** was formed in 30% yield.

12 
$$\begin{array}{r} & \underset{i \in Pr_2 \text{NEt}}{\text{Holene, reflux}} \\ 12 & \underbrace{\text{Holene, reflux}}_{i \in Pr_2 \text{NEt, PhSH}} \\ 13 & \underbrace{\text{Holene, reflux}}_{i \in Pr_2 \text{NEt, PhSH}} \\ 16 (70\%) + 13 (30\%) \end{array}$$

Scheme 4. Radical reactions of 12 and 15.

#### 2.2. Synthesis of adenine nucleoside I

When *m*-CPBA oxidation of **16** was carried out as the first step to prepare the allylic alcohol (**II** in Scheme 1), the epoxide **19** was formed as a mixture of two diastereomers (2.6:1) in 69% yield (Scheme 5, method A). NOE experiments (Fig. 1) of both products showed that the major isomer is the *endo*-**19** compound (NOE: H-5/H-6, 8.6%; H-5/H-8β, 0.5%; H-8β/H-7, 13.0%; H-8α/H-7, 1.0%), and the minor one is the *exo*-**19** compound (NOE: H-4/H-6, 3.4%; H-7/H-8α, 5.0%; H-1'/H-8β, 3.2%; H-5/H-4β, 6.3%; H-4α/H-6, 3.2%). While ring opening of the *endo*-**19** compound with phenylselenide anion generated from (PhSe)<sub>2</sub>/NaBH<sub>4</sub> resulted in poor



Scheme 5. Reagents and conditions: (a) method A: *m*-CPBA, CH<sub>2</sub>Cl<sub>2</sub> (69%, *endo/exo*=2.6:1), method B: NIS, AcOH, CH<sub>2</sub>Cl<sub>2</sub> then NaOMe, MeOH (90%, *endo/exo*=1:7); (b) (PhSe)<sub>2</sub>, NaBH<sub>4</sub>, EtOH, rt.



Figure 1. NOE experiments of the endo- and exo-19.

regioselectivity forming the selenide **20** (30%) as well as the undesired **21** (57%), the reaction of the *exo*-**19** compound gave **22** exclusively in 86% yield.

As shown in Scheme 5 by method B, the *exo*-epoxide **19** can be obtained as the major isomer upon reacting **16** with NIS/AcOH followed by NaOMe. The resulting mixture of the two epoxides, when reacted with  $(PhSe)_2/NaBH_4$ , furnished the desired selenide **22** in 83% yield from **16** after chromatographic purification.

It is interesting that both *m*-CPBA and NIS favor an approach from the more hindered concave face of **16**. Although there have been several precedents on the similar curious observation regarding electrophilic reaction in bicyclo[3.3.0] systems, <sup>14,15</sup> we have no clear explanation for the observed stereochemical outcome at the present time.

After oxidation of the selenide **22** with *m*-CPBA, selenoxide *syn*-elimination was effected in refluxing CH<sub>2</sub>Cl<sub>2</sub> in the presence of Et<sub>3</sub>N. This gave the allyl alcohol **23** in 90% yield in two steps (Scheme 6). For the introduction of the adenine base, we initially considered that the Pd-catalyzed glycosylation reported by Trost<sup>16</sup> could be employed. However, neither the acetate nor the carbonate derived from **23** gave I upon reacting with  $N^6$ -benzoyladenine/NaH in the presence of Pd(PPh<sub>3</sub>)<sub>4</sub> in DMF. We then selected the Mitsunobu reaction. Requisite inversion of the hydroxyl group of **23** was also carried out by this reaction (DEAD/PPh<sub>3</sub>/AcOH). Subsequent methanolysis gave the inverted allyl alcohol **24** in 98% yield.

As a precursor of the adenine base, 6-chloropurine was initially used in the Mitsunobu reaction of **24** (DIAD/PPh<sub>3</sub>/THF). In this step, as shown in Scheme 6, the desired **25** was accompanied by the  $N^7$ -glycoside **26** (9%). Also, ammonolysis of the crude **25** by heating in a sealed tube gave the amide **28** (5%) as a byproduct (the yield of **27**: 68% from **24**). These observations led us to examine the use of bis(Boc)adenine (**29**)<sup>17</sup> instead of 6-chloropurine.

The reaction between 24 and 29 under the Mitsunobu conditions was followed by removal of the Boc-protecting group with 50% aqueous HCO<sub>2</sub>H. Compound 27 was obtained in 83% overall yield from 24. Formation of  $N^7$ -glycosylated product was not detected in this reaction. Since 29 is more soluble in THF than 6-chloropurine, and since use of a sealed tube can be avoided, we feel this method is suitable for the preparation of carbocyclic adenine nucleosides in general. Stereochemistry of 27 was confirmed by X-ray crystallographic analysis as shown in Figure 2.

# 2.3. Conversion of the lactone nucleoside 27 to 4'-branched BCA derivatives

To prepare BCA derivatives with a variety of carbon-functionalities at the 4'-position, the lactone nucleoside **27** was first converted to the cyclic hemiacetal **30** (98%) by reacting with DIBALH. Reduction of **30** with NaBH<sub>4</sub> gave the 4'-hydroxymethyl derivative **31** in 86% yield. The Wittig reaction of **30** with PPh<sub>3</sub>=CH<sub>2</sub> proceeded well at low temperature in THF to give the vinyl derivative **32** in 96% yield. The reaction



Scheme 6. Reagents and conditions: (a) *m*-CPBA, CH<sub>2</sub>Cl<sub>2</sub> then Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, reflux (90%); (b) AcOH, DEAD, Ph<sub>3</sub>P, THF, 0 °C; (c) K<sub>2</sub>CO<sub>3</sub>, MeOH, rt (98% from **23**); (d) 6-chloropurine, DIAD, Ph<sub>3</sub>P, THF, -40 °C to rt; (e) NH<sub>3</sub>/MeOH, 120 °C in a sealed tube, **27** (68% from **24**) and **28** (5% from **24**); (f) **29**, DIAD, Ph<sub>3</sub>P, THF, -40 °C to rt; (g) aq 50% HCO<sub>2</sub>H, THF, 50 °C, **27** (83% from **24**).

with a stable ylide such as  $PPh_3$ =CHCO<sub>2</sub>Me, on the other hand, was very sluggish and required refluxing in xylene. Furthermore, the product formed in this reaction was not the expected one but the bicyclic nucleoside **33** (72% as a single stereoisomer, the stereochemistry of the 8-position was not determined), which apparently resulted from intramolecular 1,4addition of the 5'-hydroxymethyl group to the initially formed Wittig product.

The 4'-cyano derivative (**34**) was prepared from **30** in 84% yield by the following sequence of reactions: oxime formation, O-acetylation, and finally elimination of AcOH at 100 °C in the presence of NaOAc. Several attempts to synthesize the 4'-ethynyl derivative (**35**) from **30** met with failure:  $Ph_3P^+CH_2BrBr^-/tert$ -BuOK,<sup>18</sup>  $Ph_3P^+CH_2ClCl^-/LHMDS$ ,<sup>19</sup> and P(O)(OMe)<sub>2</sub>C(N<sub>2</sub>)COMe/K<sub>2</sub>CO<sub>3</sub>.<sup>20</sup> Compound **35** was formed only when **30** was reacted with TMSCHN<sub>2</sub>/LDA,<sup>21</sup> albeit in a low yield (31%) (Scheme 7).



Figure 2. ORTEP drawing of compound 27.



Scheme 7. Reagents and conditions: (a) *i*-Bu<sub>2</sub>AlH, CH<sub>2</sub>Cl<sub>2</sub>,  $-78 \degree C (98\%)$ ; (b) NaBH<sub>4</sub>, MeOH (86%); (c) Ph<sub>3</sub>PCH<sub>3</sub>Br, BuLi, THF,  $-78 \degree C$  to rt (96%); (d) Ph<sub>3</sub>P=CHCO<sub>2</sub>Me, xylene, reflux (72%); (e) NH<sub>2</sub>OH·HCl, pyridine; (f) Ac<sub>2</sub>O, *i*-Pr<sub>2</sub>NEt, MeCN; (g) NaOAc, AcOH, 100 °C (84% from **30**); (h) TMSCHN<sub>2</sub>, LDA, THF,  $-78 \degree C$  to rt (31%).

Removal of the silyl group in **31**, **32**, and **35** was performed by treatment with  $Bu_4NF$  in THF to give the corresponding free carbocyclic nucleosides **36–38** (Fig. 3). Deprotection of the cyano derivative (**34**) with  $Bu_4NF/THF$  followed by  $NH_3/MeOH$ gave the bicyclic iminoether **39** as a result of nucleophilic addition of the 5'-hydroxylmethyl group to the 4'-cyano group.

The 4'-carbon-substituted BCA analogues **36–38** did not show any activity against HIV-1 (HTLV-III<sub>B</sub> strain) replication at their non-toxic concentrations to the host cell (data not shown). We also examined their effects on genome-length HCV RNA replication using the OR6 cell-based assay system.<sup>22,23</sup> However, at 10  $\mu$ M concentration, these compounds exerted no effects on HCV RNA replication without concomitant cytotoxicity.



### 3. Conclusion

The synthesis of novel 4'-branched BCA analogues was carried out using a radical-mediated 5-exo-trig cyclization as a key step, which takes place through a  $S_H 2'$  process. The allyl sulfone 15 was found to give a higher yield of the cyclized product (16) than the allyl sulfide 14. Several experiments were carried out to see why 15 acts as a better substrate in this cyclization reaction. Transformation of 16 to the allyl alcohol 23 was effected by a series of reactions: stereoselective epoxide formation, regioselective ring opening with phenylselenide anion, and selenoxide elimination. For glycosylation under Mitsunobu conditions, the inverted allyl alcohol 24 prepared from 23 was reacted with bis(Boc)adenine (29). Manipulation of the lactone function of the resulting 27 allowed preparation of the 4'-hydroxymethyl (31), the 4'-vinyl (32), the 4'-cyano (34), and the 4'-ethynyl (35) derivatives, which were deprotected to give the corresponding free nucleosides (36-38). Deprotection of the 4'-cyano derivative (34) led to the formation of the bicyclic iminoether (39).

Compounds **36–38** were tested for their potential to inhibit replication of HIV-1 and HCV in cell culture, but no significant inhibition was observed.

#### 4. Experimental

#### 4.1. General

Melting points were determined on a Yanaco micro melting point apparatus, and are uncorrected. <sup>1</sup>H NMR and <sup>13</sup>C NMR were measured on a JEOL JNM-GX 400 (400 MHz). Chemical shifts are reported relative to Me<sub>4</sub>Si. Mass spectra (MS) were taken in FAB mode with *m*-nitrobenzyl alcohol as a matrix on a JEOL JMS-700. Infrared spectra (IR) were recorded on a JASCO FT/IR-410 spectrophotometer. Column chromatography was carried out on silica gel (Micro Bead Silica Gel PSQ 100B, Fuji Silysia Chemical Ltd). Thin layer chromatography (TLC) was performed on silica gel (precoated silica gel plate F<sub>254</sub>, Merck). Where necessary, analytical samples were purified by high-performance liquid chromatography (HPLC). HPLC was carried out on a Shimadzu LC-6AD with a Shim-pack PREP-SIL (H)\* KIT column (2×25 cm). THF was distilled from benzophenone ketyl.

## 4.2. 1-(tert-Butyldiphenylsilyloxy)methyl-4-phenylthio-2-cyclopentenecarboxylic acid methyl ester (12)

To a mixture of  $10^{10}$  (5.0 g, 12.2 mmol) and PhSH (1.5 mL, 14.6 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (60 mL) was added Et<sub>3</sub>N (68  $\mu$ L,

0.49 mmol) at room temperature. After stirring for 12 h, the reaction mixture was partitioned between 1 M HCl and  $CH_2Cl_2$ . After evaporation, the resulting **11** (6.13 g) was used for the next reaction without further purification. To a MeOH (160 mL) solution of the crude 11 (6.13 g) was added NaBH<sub>4</sub> (923 mg, 24.4 mmol) at -40 °C. The reaction mixture was stirred for 2 h. Quenching the reaction by adding acetone (40 mL)/AcOH (1.4 mL) was followed by partition between saturated aqueous NaHCO<sub>3</sub> and CH<sub>2</sub>Cl<sub>2</sub>. The organic layer was evaporated and the residue was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (65 mL). To this solution were added pyridine (1.97 mL, 24.4 mmol) and Tf<sub>2</sub>O (3.08 mL, 18.3 mmol) at -40 °C. After being stirred for 0.5 h at room temperature, the reaction mixture was partitioned between saturated aqueous NaHCO<sub>3</sub> and CH<sub>2</sub>Cl<sub>2</sub>. The organic layer was evaporated and the residue was dissolved to MeCN (65 mL). To this solution was added DBN (3.02 mL, 24.4 mmol) at 0 °C and the mixture was stirred for 0.5 h at room temperature. The mixture was partitioned between saturated aqueous NaHCO3 and CH2Cl2. Silica gel column chromatography (hexane/AcOEt=50:1) of the organic layer gave 12 (5.31 g, 87% from 10, isomeric mixture ca. 2:1) as an oil. The two isomers were separated by HPLC (hexane/AcOEt=10:1,  $t_R$  9.2 min for the minor isomer;  $t_R$ 10.9 min for the major isomer) for their NMR measurement.

### 4.2.1. NMR spectra of the major isomer

<sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 1.01 (9H, s, SiBu-*tert*), 2.04 (1H, dd, J=14.4 and 4.4 Hz, H-5), 2.94 (1H, dd, J=14.4 and 8.4 Hz, H-5), 3.68 (3H, s, OMe), 3.73 (1H, d, J=9.3 Hz, CH<sub>2</sub>OSi), 3.85 (1H, d, J=9.3 Hz, CH<sub>2</sub>OSi), 4.33–4.37 (1H, m, H-4), 5.86 (1H, dd, J=5.6 and 1.5 Hz, H-2), 5.89 (1H, dd, J=5.6 and 2.0 Hz, H-3), 7.16–7.63 (15H, m, Ph); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 19.3, 26.7, 37.6, 51.5, 52.1, 63.2, 69.3, 126.7, 127.6, 128.9, 129.6, 129.7, 131.1, 133.1, 133.2, 133.3, 134.3, 135.4, 135.6, 135.7, 178.5; FABMS *m*/*z* 503 (M<sup>+</sup>+H).

#### 4.2.2. NMR spectra of the minor isomer

<sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 1.02 (9H, s, SiBu-*tert*), 2.41–2.43 (2H, m, H-5), 3.66 (3H, s, OMe), 3.70 (1H, d, J=9.3 Hz, CH<sub>2</sub>OSi), 3.75 (1H, d, J=9.3 Hz, CH<sub>2</sub>OSi), 4.18–4.22 (1H, m, H-4), 5.88 (1H, dd, J=5.6 and 2.0 Hz, H-3), 5.94 (1H, dd, J=5.6 and 1.7 Hz, H-2), 7.20–7.63 (15H, m, Ph); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 19.3, 26.7, 37.8, 51.5, 52.0, 62.6, 69.1, 126.8, 127.7, 128.8, 129.7, 131.5, 133.1, 133.2, 133.3, 134.0, 135.2, 135.5, 135.8, 174.5; FABMS *m/z* 503 (M<sup>+</sup>+H). Anal. Calcd for C<sub>30</sub>H<sub>34</sub>O<sub>3</sub>SSi: C, 71.67; H, 6.82. Found: C, 71.38; H, 6.86.

## 4.3. Benzenesulfonyl-1-(tert-butyldiphenylsilyloxy)methyl-2-cyclopentenecarboxylic acid methyl ester (13)

To a CH<sub>2</sub>Cl<sub>2</sub> (30 mL) solution of **12** (865 mg, 1.72 mmol) was added *m*-CPBA (>65%, 1.0 g, 3.78 mmol). The reaction mixture was stirred for 3 h at room temperature, treated with Et<sub>3</sub>N (527  $\mu$ L, 3.78 mmol), and then partitioned between saturated aqueous NaHCO<sub>3</sub> and CH<sub>2</sub>Cl<sub>2</sub>. Silica gel column chromatography (hexane/AcOEt=5:1) of the organic layer gave

**13** (879 mg, 96%, inseparable mixture of two isomers ca. 2:1) as an oil.

#### 4.3.1. NMR spectra of the major isomer

<sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.02 (9H, s, SiBu-*tert*), 2.26 (1H, dd, J=15.0 and 5.2 Hz, H-5), 2.82 (1H, dd, J=15.0 and 9.5 Hz, H-5), 3.62 (1H, d, J=9.5 Hz, CH<sub>2</sub>OSi), 3.65 (3H, s, OMe), 3.77 (1H, d, J=9.5 Hz, CH<sub>2</sub>OSi), 4.41–4.44 (1H, m, H-4), 5.83 (1H, dd, J=5.7 and 1.7 Hz, H-3), 6.06 (1H, dd, J=5.7 and 2.3 Hz, H-2), 7.37–7.47 (8H, m, Ph), 7.54–7.62 (5H, m, Ph), 7.80–7.82 (2H, m, Ph); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  19.3, 26.6, 30.5, 52.3, 63.7, 69.1, 71.6, 126.6, 127.6, 127.7, 128.9, 129.1, 129.7, 129.8, 133.1, 133.8, 135.5, 135.6, 137.3, 138.6, 173.6.

## 4.3.2. NMR spectra of the minor isomer

<sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 0.99 (9H, s, SiBu-*tert*), 2.22 (1H, dd, J=15.0 and 9.2 Hz, H-5), 2.69 (1H, dd, J=15.0 and 5.2 Hz, H-5), 3.56 (3H, s, OMe), 3.62 (1H, d, J=9.2 Hz, CH<sub>2</sub>OSi), 3.68 (1H, d, J=9.2 Hz, CH<sub>2</sub>OSi), 4.27–4.29 (1H, m, H-4), 5.85 (1H, dd, J=5.7 and 2.3 Hz, H-3), 6.15 (1H, dd, J=5.7 and 2.3 Hz, H-2), 7.34–7.37 (4H, m, Ph), 7.40–7.43 (2H, m, Ph), 7.54–7.57 (6H, m, Ph), 7.63–7.66 (1H, m, Ph), 7.85–7.87 (2H, m, Ph); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 19.2, 26.6, 30.4, 52.2, 62.9, 69.0, 71.2, 126.1, 127.7, 129.0, 129.3, 129.8, 132.7, 132.8, 133.7, 135.4, 135.5, 136.7, 139.5, 173.1; FABMS *m*/*z* 535 (M<sup>+</sup>+H). Anal. Calcd for C<sub>30</sub>H<sub>34</sub>O<sub>5</sub>SSi: C, 67.38; H, 6.41. Found: C, 67.39; H, 6.54.

## 4.4. 1-(tert-Butyldiphenylsilyloxy)methyl-4-phenylthio-2-cyclopentenecarboxylic acid (phenylseleno)methyl ester (14)

To a mixture of 12 (4.01 g, 7.98 mmol), THF (60 mL), and MeOH (90 mL) was added aqueous KOH [KOH (2.24 g, 39.9 mmol)/H<sub>2</sub>O (30 mL)]. After being stirred at room temperature for 2 days, the reaction mixture was partition between 1 M HCl and CH<sub>2</sub>Cl<sub>2</sub>. The organic layer was evaporated and the residue was dissolved in DME (26 mL), to which PhSeCH<sub>2</sub>Cl (1.9 g, 9.24 mmol), NaI (1.39 g, 9.24 mmol), and *i*-Pr<sub>2</sub>NEt (1.6 mL, 9.24 mmol) were added. The whole mixture was refluxed for 24 h and partitioned between NaHCO<sub>3</sub> and Et<sub>2</sub>O. Flash silica gel column chromatography (hexane/AcOEt=80:1) of the organic layer gave 14 (3.57 g, 68% from 12, a mixture of two isomers ca. 10:4) as an oil. Compound 14 was used for the radical reaction without further purification due to its instability to silica gel. <sup>1</sup>H NMR (CDCl<sub>3</sub>) & 1.01 (12.6H, s, SiBu-tert), 2.01 (1H, dd, J=14.4 and 4.6 Hz, H-5), 2.40-2.42 (0.8H, m, H-5×2), 2.89 (1H, dd, J=14.4 and 8.3 Hz, H-5), 3.71 (0.4H, d, J=9.5 Hz, CH<sub>2</sub>OSi), 3.75 (1H, d, J=9.5 Hz, CH<sub>2</sub>OSi), 3.76 (0.4H, d, J=9.5 Hz, CH<sub>2</sub>OSi), 3.86 (1H, d, J=9.5 Hz, CH<sub>2</sub>OSi), 4.18-4.23 (0.4H, m, H-4), 4.29-4.33 (1H, m, H-4), 5.49-5.61 (2.8H, m, CH<sub>2</sub>Se×4), 5.81 (1H, dd, J=5.4 and 1.7 Hz, H-2), 5.88-5.90 (1.8H, m, H-2 and H-3), 7.16-7.63 (20H, m, Ph);  ${}^{13}$ C NMR (CDCl<sub>3</sub>)  $\delta$  19.3, 26.7, 37.5, 37.7, 51.5, 62.5, 62.6, 62.7, 63.3, 68.6, 68.9, 126.8, 127.6, 127.7, 128.9, 129.2, 129.7, 129.8, 131.1, 131.4, 132.5, 132.8, 133.0, 133.1, 133.2, 133.3, 134.5, 134.9, 135.3, 135.6, 135.7, 173.2, 173.3; FABMS *m*/*z* 697 (M<sup>+</sup>+H).

## 4.5. 4-Benzenesulfonyl-1-(tert-butyldiphenylsilyloxy)methyl-2-cyclopentenecarboxylic acid (phenylseleno)methyl ester (15)

Compound 13 (9.95 g, 18.6 mmol) was treated by the procedure described for the preparation of 14. This gave 15 (10.4 g, 81% from 13, a mixture of two isomers ca. 10:7) as an oil. Compound 15 was used for the radical reaction without further purification due to its instability to silica gel. <sup>1</sup>H NMR (CDCl<sub>3</sub>) & 0.98 (9H, s, SiBu-tert), 1.00 (6.3H, s, SiBu-tert), 2.21–2.27 (1.7H, m, H-5×2), 2.67 (1H, dd, J=15.0 and 5.4 Hz, H-5), 2.75 (0.7H, dd, J=15.0 and 9.3 Hz, H-5), 3.62-3.80 (3.4H, m, CH<sub>2</sub>OSi×4), 5.38-5.59 (3.4H, m, CH<sub>2</sub>Se×4), 5.82 (0.7H, dd, J=5.6 and 2.2 Hz, H-2 or H-3), 5.86 (1H, dd, J=5.6 and 2.2 Hz, H-2 or H-3), 6.00 (0.7H, dd, J=5.6 and 2.2 Hz, H-2 or H-3), 6.06 (1H, dd, J=5.6 and 2.2 Hz, H-2 or H-3), 7.22–7.85 (34H, m, Ph);  $^{13}$ C NMR (CDCl<sub>3</sub>)  $\delta$  19.3, 19.4, 26.7, 30.2, 30.4, 62.7, 62.8, 63.0, 63.8, 68.4, 68.7, 71.3, 71.5, 126.8, 127.2, 127.7, 127.8, 128.9, 129.0, 129.1, 129.2, 129.3, 129.7, 129.8, 129.9, 132.6, 132.7, 133.0, 133.1, 133.2, 133.8, 133.9, 135.5, 135.6, 135.7, 137.3, 138.1, 138.8, 171.7, 172.4; FABMS *m*/*z* 729 (M<sup>+</sup>+K).

#### 4.6. Radical reaction of 14

To a refluxing toluene (190 mL) solution of **14** (4.99 g, 7.58 mmol) and *i*-Pr<sub>2</sub>NEt (2.18 mL, 12.5 mmol) was added dropwise a toluene (20 mL) solution of Bu<sub>3</sub>SnH (4.77 mL, 1.77 mmol) and AIBN (245 mg, 1.52 mmol) over 5 h under positive pressure of dry Ar. After evaporation of the solvent, the reaction mixture was purified by silica gel chromatography (hexane/AcOEt=30:1). This gave **16** (1.96 g, 61% as a solid), **17** (150 mg, 5% as an oil), and **18** (538 mg, 18% as an oil).

#### 4.6.1. Physical data for 16

Mp 77–78 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.05 (9H, s, SiBu-*tert*), 2.20–2.26 (1H, m, CH<sub>2</sub>CH=CH), 2.65–2.71 (1H, m, CH<sub>2</sub>CH=CH), 3.57–3.60 (1H, m, H-5), 3.66 (1H, d, J=9.6 Hz, CH<sub>2</sub>OSi), 4.11(1H, d, J=9.6 Hz, CH<sub>2</sub>OSi), 4.19 (1H, dd, J=8.7 and 1.7 Hz, CH<sub>2</sub>OCO), 4.56 (1H, dd, J=8.7 and 7.6 Hz, CH<sub>2</sub>OCO), 5.63–5.65 and 5.72–5.75 (2H, each as m, CH=CH), 7.36–7.47 (6H, m, Ph), 7.62–7.73 (4H, m, Ph); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  19.2, 26.7, 39.3, 50.3, 56.7, 67.3, 71.8, 127.9, 129.9, 130.0, 131.0, 132.2, 133.1, 135.5 135.7, 181.9; FABMS *m*/*z* 431 (M<sup>+</sup>+K). Anal. Calcd for C<sub>24</sub>H<sub>28</sub>O<sub>3</sub>Si: C, 73.43; H, 7.19. Found: C, 73.39; H, 7.29.

## 4.6.2. Physical data for 17

<sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 1.04 (9H, s, SiBu-*tert*), 1.40–1.48 (1H, m,  $-CH_2CH_2-$ ), 1.56–1.70 (3H, m,  $-CH_2CH_2-$ ), 1.89–2.00 (2H, m,  $-CH_2CH_2-$ ), 2.91–2.93 (1H, m, H-5), 3.50 (1H, d, *J*=9.5 Hz, CH<sub>2</sub>OSi), 3.99 (1H, dd, *J*=9.0 and 3.2 Hz, CH<sub>2</sub>OCO), 4.05 (1H, d, *J*=9.5 Hz, CH<sub>2</sub>OSi), 4.54 (1H, t, *J*=9.0 Hz, CH<sub>2</sub>OCO), 7.37–7.47 (6H, m, Ph), 7.63–7.65 (4H, m, Ph); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  19.2, 25.5, 26.7, 33.7, 34.4, 42.7, 58.5, 67.5, 73.3, 127.8, 129.8, 129.9, 132.5, 133.2, 135.5, 135.7, 181.8; FABMS *m*/*z* 395 (M<sup>+</sup>+H). Anal. Calcd for C<sub>24</sub>H<sub>30</sub>O<sub>3</sub>Si: C, 73.05; H, 7.66. Found: C, 72.78; H, 7.68.

## 4.6.3. Physical data for 18

<sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 1.05 (9H, s, SiBu-*tert*), 1.89–1.95 (1H, m,  $-CH_2CH_2-$ ), 2.31–2.43 (3H, m,  $-CH_2CH_2-$ ), 3.69 (3H, s, OCH<sub>3</sub>), 3.72 (1H, d, *J*=9.3 Hz, CH<sub>2</sub>OSi), 3.81 (1H, d, *J*= 9.3 Hz, CH<sub>2</sub>OSi), 5.74–5.76 and 5.86–5.88 (2H, each as m, -CH=CH-), 7.36–7.44 (6H, m, Ph), 7.62–7.65 (4H, m, Ph); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 18.9, 26.3, 29.6, 31.2, 51.5, 62.6, 68.6, 127.2, 129.2, 130.7, 133.1, 133.5, 135.2, 135.3, 175.2; FABMS *m*/*z* 395 (M<sup>+</sup>+H). Anal. Calcd for C<sub>24</sub>H<sub>30</sub>O<sub>3</sub>Si: C, 73.05; H, 7.66. Found: C, 72.99; H, 7.87.

#### 4.7. Radical reaction of 15

Compound **15** (4.50 g, 6.52 mmol) was treated by the procedure described for the reaction of **14**. This gave **16** (2.40 g, 94%).

### 4.8. Radical reaction of 12 (depicted in Scheme 4)

Compound 12 (99 mg, 0.917 mmol) was treated by the procedure described for the reaction of 14. After evaporation of the solvent, the reaction mixture was analyzed by <sup>1</sup>H NMR (12/18=85:15, calculated by integrating OMe).

#### 4.9. Radical reaction of 15 (depicted in Scheme 4)

To a refluxing toluene (5 mL) solution of **15** (138 mg, 0.20 mmol), *i*-Pr<sub>2</sub>NEt (105  $\mu$ L, 0.60 mmol), and PhSH (10.3  $\mu$ L, 0.10 mmol) was added dropwise a toluene (1 mL) solution of Bu<sub>3</sub>SnH (108  $\mu$ L, 0.40 mmol) and AIBN (6.6 mg, 0.04 mmol) over 5 h under positive pressure of dry Ar. After evaporation of the solvent, the reaction mixture was analyzed by <sup>1</sup>H NMR (**16/13**=70:30, calculated by integrating CH<sub>2</sub>OSi).

#### 4.10. Preparation of the epoxide 19

#### 4.10.1. Method A

To a CH<sub>2</sub>Cl<sub>2</sub> (5 mL) solution of **16** (341 mg, 0.87 mmol) was added *m*-CPBA (>65%, 242 mg, 0.91 mmol) and the mixture was stirred for 17 h at room temperature. The reaction mixture was treated with Et<sub>3</sub>N (0.127 mL, 0.91 mmol) and then partitioned between saturated aqueous NaHCO<sub>3</sub> and CH<sub>2</sub>Cl<sub>2</sub>. Preparative TLC (hexane/AcOEt=2:1) of the organic layer gave *endo*-**19** (176 mg, 50%, foam) and *exo*-**19** (69 mg, 19%, foam).

4.10.1.1. Physical data for endo-**19**. <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.04 (9H, s, SiBu-*tert*), 1.70 (1H, dd, *J*=14.6 and 2.2 Hz, H-8), 2.45 (1H, d, *J*=14.6 Hz, H-8), 2.81–2.84 (1H, m, H-5), 3.54–3.57 (2H, m, CH<sub>2</sub>OSi and H-6), 3.65 (1H, t, *J*=2.2 Hz, H-7), 3.91 (1H, d, *J*=9.8 Hz, CH<sub>2</sub>OSi), 4.48 (1H, dd, *J*=9.0 and

7.1 Hz, H-4), 4.53 (1H, dd, J=9.0 and 1.7 Hz, H-4), 7.37–7.48 (6H, m, Ph), 7.59–7.63 (4H, m, Ph); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  19.2, 26.7, 34.3, 44.3, 54.9, 58.3, 60.4, 66.6, 68.4, 127.9, 130.0, 130.1, 135.4, 135.7, 180.0; FABMS *m*/*z* 409 (M<sup>+</sup>+H). Anal. Calcd for C<sub>24</sub>H<sub>28</sub>O<sub>4</sub>Si: C, 70.55; H, 6.91. Found: C, 70.22; H, 6.89.

4.10.1.2. Physical data for exo-19. <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.02 (9H, s, SiBu-tert), 1.82 (1H, d, J=15.0 Hz, H-8), 2.16 (1H, dd, J=15.0 and 2.2 Hz, H-8), 3.45 (1H, dd, J=9.5 and 5.6 Hz, H-5), 3.51 (1H, d, J=9.3 Hz, CH<sub>2</sub>OSi), 3.52 (1H, d, J=2.2 Hz, H-6), 3.56 (1H, t, J=2.2 Hz, H-7), 3.96 (1H, d, J=9.3 Hz, CH<sub>2</sub>OSi), 4.15 (1H, dd, J=9.5 and 5.6 Hz, H-4), 4.40 (1H, t, J=9.5 Hz, H-4), 7.37–7.46 (6H, m, Ph), 7.61–7.64 (4H, m, Ph); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  18.7, 26.2, 34.1, 43.9, 57.3, 57.6, 61.4, 66.3, 67.2, 127.3, 127.4, 129.4, 131.9, 132.5, 135.0, 135.2, 179.7; FABMS *m*/*z* 409 (M<sup>+</sup>+H). Anal. Calcd for C<sub>24</sub>H<sub>28</sub>O<sub>4</sub>Si: C, 70.55; H, 6.91. Found: C, 70.40; H, 6.93.

#### 4.10.2. Method B

To a mixture of **16** (837 mg, 2.13 mmol) and AcOH (1.23 mL, 21.3 mmol) in  $CH_2Cl_2$  (20 mL) wad added NIS (1.92 g, 8.52 mmol). The resulting mixture was stirred for 87 h at room temperature in the dark. After addition of saturated aqueous Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> (50 mL), the reaction mixture was partitioned between saturated aqueous NaHCO<sub>3</sub> and CH<sub>2</sub>Cl<sub>2</sub>. The organic layer was evaporated and treated with NaOMe (345 mg, 6.39 mmol) in MeOH (18 mL) for 30 min at room temperature. The reaction mixture was partitioned between 0.5 M HCl and CH<sub>2</sub>Cl<sub>2</sub>. Silica gel column chromatography (hexane/AcOEt=4:1) of the organic layer gave **19** (785 mg, 90%, *exolendo*=7:1, calculated by integrating CH<sub>2</sub>OSi). Compound **19** was used for the next ring opening reaction without separation.

## 4.11. Epoxide ring cleavage of endo-**19** by phenylselenide anion

The phenylselenide anion was prepared from  $(PhSe)_2$  (131 mg, 0.41 mmol)/EtOH (10 mL) and NaBH<sub>4</sub> (31 mg, 0.83 mmol) with stirring for 10 min at room temperature. To this solution, an EtOH (3 mL) solution of *endo*-**19** (215 mg, 0.52 mmol) was added. After being stirred for 5 h at room temperature, the reaction mixture was partitioned between 0.5 M HCl and CH<sub>2</sub>Cl<sub>2</sub>. Preparative TLC (hexane/AcOEt= 2:1) of the organic layer gave **20** (87 mg, 30%, foam) and **21** (169 mg, 57%, solid).

#### 4.11.1. Physical data for 20

<sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 1.02 (9H, s, SiBu-*tert*), 1.60 (1H, dd, J=13.6 and 10.9 Hz, H-8), 2.31 (1H, br, OH), 2.38 (1H, dd, J=13.6 and 6.6 Hz, H-8), 3.14–3.18 (1H, m, H-5), 3.22–3.27 (1H, m, H-7), 3.52 (1H, d, J=9.7 Hz, CH<sub>2</sub>OSi), 3.93 (1H, d, J=9.7 Hz, CH<sub>2</sub>OSi), 4.05 (1H, t, J=8.0 Hz, H-6), 4.31 (1H, t, J=9.7 Hz, H-4), 4.62 (1H, dd, J=9.7 and 3.4 Hz, H-4), 7.24–7.32 (3H, m, Ph), 7.37–7.46 (6H, m, Ph), 7.50–7.52 (2H, m, Ph), 7.59–7.63 (4H, m, Ph); <sup>13</sup>C NMR (CDCl<sub>3</sub>)

δ 19.2, 26.7, 36.6, 44.6, 46.5, 55.8, 65.8, 67.0, 77.5, 126.6, 127.8, 128.4, 129.4, 129.9, 132.2, 132.9, 135.3, 135.5, 135.6, 180.8; FABMS *m*/*z* 605 (M<sup>+</sup>+K). Anal. Calcd for C<sub>30</sub>H<sub>34</sub>O<sub>4</sub>SeSi: C, 63.70; H, 6.06. Found: C, 63.33; H, 6.04.

#### 4.11.2. Physical data for 21

Mp 127–128 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 1.02 (9H, s, SiButert), 1.99–2.09 (2H, m, H-8), 2.27 (1H, br, OH), 2.90–2.93 (1H, m, H-5), 3.45–3.47 (2H, m, H-6 and CH<sub>2</sub>OSi), 4.00 (1H, d, *J*=9.2 Hz, CH<sub>2</sub>OSi), 4.12–4.15 (1H, m, H-7), 4.27 (1H, dd, *J*=9.2 and 2.9 Hz, H-4), 4.50 (1H, t, *J*=9.2 Hz, H-4), 7.25–7.32 (3H, m, Ph), 7.35–7.53 (6H, m, Ph), 7.58–7.59 (2H, m, Ph), 7.60–7.62 (4H, m, Ph); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 19.2, 26.7, 39.3, 47.8, 54.5, 56.3, 68.0, 72.7, 77.3, 127.8, 127.9, 128.3, 129.4, 129.9, 132.2, 132.8, 134.7, 135.5, 136.6, 181.5; FABMS *m*/*z* 567 (M<sup>+</sup>+H). Anal. Calcd for C<sub>30</sub>H<sub>34</sub>O<sub>4</sub>SeSi: C, 63.70; H, 6.06. Found: C, 63.44; H, 6.08.

## 4.12. Epoxide ring cleavage of exo-**19** by phenylselenide anion: formation of the allyl alcohol **22**

Compound **22** was prepared from *exo*-**19** (110 mg, 0.22 mmol) by the procedure described for the reaction of *endo*-**19**. Silica gel column chromatography (CH<sub>2</sub>Cl<sub>2</sub>) gave **22** (131 mg, 86%) as a foam.

Compound **22** was also obtained in 83% yield from **16** through method B in Scheme 5 without isolating *exo*-**19**.

#### 4.12.1. Physical data for 22

<sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 1.02 (9H, s, SiBu-*tert*), 1.93 (1H, dd, J=14.4 and 10.2 Hz, H-8), 2.08 (1H, dd, J=14.4 and 8.1 Hz, H-8), 2.33 (1H, d, J=3.2 Hz, OH), 2.78–2.82 (1H, m, H-5), 3.22–3.29 (1H, m, H-7), 3.57 (1H, d, J=9.5 Hz, CH<sub>2</sub>OSi), 3.80–3.85 (1H, m, H-6), 3.94 (1H, d, J=9.5 Hz, CH<sub>2</sub>OSi), 4.33 (1H, dd, J=9.3 and 1.9 Hz, H-4), 4.50 (1H, dd, J=9.3 and 7.3 Hz, H-4), 7.26–7.63 (15H, m, Ph); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 18.9, 26.4, 35.7, 47.7, 49.9, 53.5, 67.7, 70.2, 81.7, 127.6, 127.7, 128.3, 129.2, 129.8, 131.7, 132.4, 135.2, 135.3, 135.4, 180.4; FABMS m/z 605 (M<sup>+</sup>+K). Anal. Calcd for C<sub>30</sub>H<sub>34</sub>O<sub>4</sub>SeSi: C, 63.70; H, 6.06. Found: C, 63.54; H, 6.14.

## 4.13. $(\pm)$ -c-1-(tert-Butyldiphenylsilyloxy)methyl-4hydroxy-7-oxabicyclo-[3.3.0]oct-2-en-8-one (23)

To a CH<sub>2</sub>Cl<sub>2</sub> (50 mL) solution of **22** (1.9 g, 3.36 mmol) was added *m*-CPBA (>65%, 1.07 g, 4.03 mmol) and the mixture was stirred for 0.5 h at room temperature. After addition of Et<sub>3</sub>N (2.34 mL, 16.8 mmol), the reaction mixture was refluxed for 1 h and then partitioned between saturated aqueous NaHCO<sub>3</sub> and CH<sub>2</sub>Cl<sub>2</sub>. Silica gel column chromatography (hexane/AcOEt=1:2) of the organic layer gave **23** (1.23 g, 90%, solid). Mp 120–121 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.06 (9H, s, SiBu-*tert*), 2.43 (1H, d, *J*=10.3 Hz, OH), 3.04 (1H, dd, *J*=9.5 and 6.9 Hz, H-5), 3.82 (1H, d, *J*=10.3 Hz, CH<sub>2</sub>OSi), 3.89 (1H, dd, *J*=9.5 and 6.9 Hz, H-4), 3.91 (1H, d, *J*= 10.3 Hz, CH<sub>2</sub>OSi), 4.54 (1H, dd, *J*=10.3 and 2.3 Hz, H-6), 4.61 (1H, t, *J*=9.5 Hz, H-4), 5.73 (1H, d, *J*=5.2 Hz, H-8),

6.12 (1H, dd, J=5.2 and 2.3 Hz, H-7), 7.38–7.47 (6H, m, Ph), 7.61–7.64 (4H, m, Ph); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  19.2, 26.8, 51.5, 63.5, 65.1, 69.6, 81.3, 127.9, 130.1, 131.9, 132.0, 132.1, 135.6, 136.3, 175.8; FABMS *m*/*z* 409 (M<sup>+</sup>+H). Anal. Calcd for C<sub>24</sub>H<sub>28</sub>O<sub>4</sub>Si: C, 70.55; H, 6.91. Found: C, 70.41; H, 6.95.

## 4.14. (±)-t-1-(tert-Butyldiphenylsilyloxy)methyl-4-hydroxy-7-oxabicyclo-[3.3.0]oct-2-en-8-one (24)

To a stirred THF (2 mL) solution containing 23 (105 mg. 0.257 mmol), PPh<sub>3</sub> (202 mg, 0.771 mmol), and AcOH (73.6 µL, 1.29 mmol) was added DEAD (2.2 M in toluene, 352 µL, 0.771 mmol) at 0 °C. After stirring for 15 min, the reaction mixture was partitioned between saturated aqueous NaHCO<sub>3</sub> and CH<sub>2</sub>Cl<sub>2</sub>. The organic layer was evaporated and the resulting residue was dissolved in MeOH (4 mL) containing K<sub>2</sub>CO<sub>3</sub> (80 mg, 0.514 mmol). After being stirred for 15 min at 0 °C, the reaction mixture was partitioned between 0.5 M HCl and CH<sub>2</sub>Cl<sub>2</sub>. Silica gel column chromatography (hexane/AcOEt=3:1) of the organic layer gave 24 (103 mg, 98%, solid). Mp 139–140 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 1.05 (9H, s, SiBu-tert), 1.68 (1H, d, J=7.3 Hz, OH), 3.21-3.26 (1H, m, H-5), 3.55 (1H, d, J=9.8 Hz, CH<sub>2</sub>OSi), 4.12 (1H, d, J=9.8 Hz, CH<sub>2</sub>OSi), 4.40 (1H, t, J=9.0 Hz, H-4), 4.64 (1H, dd, J=9.0 and 4.9 Hz, H-4), 5.00-5.04 (1H, m H-6), 5.67 (1H, dd, J=5.6 and 1.5 Hz, H-8), 5.94 (1H, dd, J=5.6 and 2.0 Hz, H-7), 7.37–7.47 (6H, m, Ph), 7.62–7.66 (4H, m, Ph); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 19.0, 26.5, 44.6, 64.0, 65.6, 65.8, 76.9, 127.6, 129.7, 130.8, 132.1, 132.7, 135.3, 135.4, 136.9, 177.5; FABMS m/z 409 (M<sup>+</sup>+H). Anal. Calcd for C<sub>24</sub>H<sub>28</sub>O<sub>4</sub>Si: C, 70.55; H, 6.91. Found: C, 70.45; H, 6.98.

## 4.15. Mitsunobu reaction between **24** and 6-chloropurine, and subsequent ammonolysis

To a stirred THF (100 mL) solution containing 24 (2.0 g, 4.90 mmol), PPh<sub>3</sub> (3.91 g, 12.2 mmol), and 6-chloropurine (1.13 g, 7.34 mmol) was added dropwise DIAD (6.42 mL, 12.2 mmol) at 0 °C. After being stirred for 1 h at 0 °C, the reaction mixture was partitioned between saturated aqueous NaHCO<sub>3</sub> and CH<sub>2</sub>Cl<sub>2</sub>. The organic layer was purified by silica gel column chromatography. Elution with hexane/AcOEt=3:1 gave a mixture of 25 and triphenylphosphine oxide. Elution with hexane/AcOEt=1:1 gave 26 (240 mg, 9%, foam). The crude 25 dissolved in THF (10 mL) was transferred in a sealed tube containing NH<sub>3</sub>/MeOH (200 mL) and the whole mixture was heated at 120 °C for 6 h. After evaporation, the reaction mixture was purified by silica gel column chromatography. Elution with CHCl<sub>3</sub>/MeOH=80:1 gave 27 (1.75 g, 68% from 24, solid), which was recrystallized from MeOH/CH<sub>2</sub>Cl<sub>2</sub>. Elution with CHCl<sub>3</sub>/MeOH=30:1 gave 28 (120 mg, 5% from 24, solid).

## 4.15.1. Physical data for 26

<sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  0.97 (9H, s, SiBu-*tert*), 3.02–3.03 (1H, m H-5'), 3.48 (1H, d, *J*=10.0 Hz, CH<sub>2</sub>OSi), 4.38 (1H, d,

J=10.0 Hz, CH<sub>2</sub>OSi), 4.41 (1H, dd, J=10.0 and 5.2 Hz, H-4'), 4.76 (1H, t, J=10.0 Hz, H-4'), 5.92−5.93 (1H, m, H-6'), 6.11 (1H, dd, J=5.2 and 2.3 Hz, H-8'), 6.23 (1H, dd, J=5.2 and 1.7 Hz, H-7'), 7.12−7.20 (3H, m, Ph), 7.28−7.31 (2H, m, Ph), 7.34−7.38 (3H, m, Ph), 7.49−7.51 (2H, m, Ph), 7.80 (1H, s, H-8), 8.95 (1H, s, H-2); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  19.0, 26.6, 49.8, 65.5, 68.8, 70.2, 121.8, 127.7, 127.8, 129.5, 129.9, 130.0, 131.8, 132.2, 135.0, 135.4, 137.2, 142.5, 145.6, 152.6, 162.3, 174.6; FABMS *m*/*z* 545 (M<sup>+</sup>+H). Anal. Calcd for C<sub>29</sub>H<sub>29</sub>ClN<sub>4</sub>O<sub>3</sub>Si: C, 63.90; H, 5.36; N, 10.28. Found: C, 63.62; H, 5.32; N, 10.21.

#### 4.15.2. Physical data for 27

Mp 229–231 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  0.99 (9H, s, SiButert), 3.13–3.15 (1H, m, H-5'), 3.72 (1H, d, *J*=10.0 Hz, CH<sub>2</sub>OSi), 4.32 (1H, d, *J*=10.0 Hz, CH<sub>2</sub>OSi), 4.58 (1H, dd, *J*=9.7 and 3.4 Hz, H-4'), 4.70 (1H, t, *J*=9.7 Hz, H-4'), 5.89 (2H, br, NH<sub>2</sub>), 6.02 (1H, dd, *J*=5.4 and 2.0 Hz, H-8'), 6.05 (1H, dd, *J*=5.4 and 2.0 Hz, H-7'), 6.59–6.60 (1H, m, H-6'), 7.25–7.58 (10H, m, Ph), 7.59 (1H, s, H-8), 8.33 (1H, s, H-2); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  19.1, 26.7, 50.3, 65.2, 65.9, 67.1, 71.7, 120.1, 127.8, 127.9, 129.9, 130.0, 130.7, 131.9, 132.5, 135.0, 135.3, 135.6, 137.0, 149.7, 153.2, 155.5, 175.9; FABMS *m*/*z* 526 (M<sup>+</sup>+H). Anal. Calcd for C<sub>29</sub>H<sub>31</sub>N<sub>5</sub>O<sub>3</sub>Si: C, 66.26; H, 5.94; N, 13.32. Found: C, 66.06; H, 5.85; N, 13.44.

#### 4.15.3. Physical data for 28

Mp 200–202 °C; <sup>1</sup>H NMR (DMSO- $d_6$ ) δ 1.00 (9H, s, SiButert), 2.64 (1H, dd, J=14.5 and 7.2 Hz, H-5'), 3.53–3.59 (1H, m, CH<sub>2</sub>OH), 3.68–3.74 (1H, m, CH<sub>2</sub>OH), 3.90 (1H, d, J=9.7 Hz, CH<sub>2</sub>OSi), 4.11 (1H, d, J=9.7 Hz, CH<sub>2</sub>OSi), 4.54 (1H, dd, J=5.7 and 4.5 Hz, OH), 5.52–5.55 (1H, m, H-1'), 6.10 (1H, dd, J=5.7 and 1.8 Hz, CH=CH), 6.22 (1H, dd, J=5.7 and 2.0 Hz, CH=CH), 6.98 (1H, br, NH<sub>2</sub>), 7.22 (2H, br, NH<sub>2</sub>), 7.34 (1H, br, NH<sub>2</sub>), 7.40–7.48 (6H, m, Ph), 7.64–7.68 (4H, m, Ph), 7.94 (1H, s, H-8), 8.06 (1H, s, H-2); <sup>13</sup>C NMR (DMSO- $d_6$ ) δ 19.0, 26.6, 52.4, 60.7, 62.7, 63.2, 67.4, 79.2, 119.0, 127.8, 127.9, 129.8, 132.8, 132.9, 135.1, 135.2, 136.5, 138.9, 149.4, 152.2, 156.0, 173.0; FABMS *m*/*z* 543 (M<sup>+</sup>+H). Anal. Calcd for C<sub>29</sub>H<sub>34</sub>N<sub>6</sub>O<sub>3</sub>Si·H<sub>2</sub>O: C, 62.12; H, 6.47; N, 14.99. Found: C, 62.39; H, 6.12; N, 14.91.

## 4.16. Mitsunobu reaction between 24 and bis(Boc)adenine (29), and subsequent hydrolysis

To a THF (50 mL) solution containing **24** (2.89 g, 7.07 mmol), Ph<sub>3</sub>P (2.36 g, 8.98 mmol), and bis(Boc)adenine (**29**)<sup>18</sup> (3.01 g, 8.98 mmol) was added dropwise DIAD (1.77 mL, 8.98 mmol) at -40 °C. The resulting mixture was stirred for 18 h at -40 °C. After evaporation of the solvent, the reaction mixture was purified by flash silica gel column chromatography (hexane/AcOEt=1:1). The crude product obtained was dissolved in THF (25 mL) containing 50% aqueous HCO<sub>2</sub>H (50 mL) and the whole mixture was heated at 50 °C for 9 h. After evaporation, the residue was treated with 26% NH<sub>4</sub>OH (2 mL) and then evaporated to dryness. Silica gel column chromatography (CHCl<sub>3</sub>/MeOH=11:1) of the residue gave **27** (3.09 g, 83%).

## 4.17. $(\pm)$ -c-4-(Adenine-9-yl)-1-(tert-butyldiphenylsilyloxy)methyl-7-oxabicyclo-[3.3.0]oct-2-en-8-ol: the cyclic hemiacetal **30**

To a CH<sub>2</sub>Cl<sub>2</sub> (10 mL) solution of **27** (200 mg, 0.49 mmol) was added *i*-Bu<sub>2</sub>AlH (1.01 M in toluene, 1.46 mL, 1.47 mmol) at -70 °C. The resulting mixture was stirred for 15 min at -70 °C and the reaction was quenched by adding saturated aqueous NH<sub>4</sub>Cl. The reaction mixture was partitioned between 0.5 M HCl and CH<sub>2</sub>Cl<sub>2</sub>. Silica gel column chromatography (CHCl<sub>3</sub>/MeOH=10:1) of the organic layer gave **30** (194 mg, 97%, a mixture of two isomers ca. 4:1) as a foam.

#### 4.17.1. Physical data for 30

<sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  0.96 and 0.97 (11.25H, each as s, SiBu-tert), 2.60-2.64 (1H, m, H-5'), 3.68 (1H, d, J=10.0 Hz, CH2OSi), 3.74-3.75 (0.5H, m, CH2OSi), 3.93-4.02 (2.5H, m, CH<sub>2</sub>OSi and H-4'), 5.19 (1H, d, J=4.4 Hz, OCHOH), 5.24 (0.25H, d, J=5.0 Hz, OCHOH), 5.35-5.37 (1.25H, m, H-6'), 5.97 (0.25H, dd, J=5.6 and 2.0 Hz, CH=CH), 6.01 (0.25H, dd, J=5.6 and 1.7 Hz, CH=CH), 6.11 (1H, dd, J=5.6 and 2.2 Hz, CH=CH), 6.30 (1H, dd, J=5.6 and 1.7 Hz, CH=CH), 6.33 (1H, d, J=4.4 Hz, OH), 6.50 (0.25H, d, J=5.0 Hz, OH), 7.20 (2.5H, br, NH<sub>2</sub>), 7.33-7.64 (12.5H, m, Ph), 7.75 (1H, s, H-8), 7.82 (0.25H, s, H-8), 8.07 (0.25H, s, H-2), 8.10 (1H, s, H-2); <sup>13</sup>C NMR (DMSO- $d_6$ )  $\delta$  18.9, 26.6, 26.7, 51.7, 52.5, 64.8, 66.0, 66.3, 67.9, 68.0, 68.4, 70.5, 99.1, 99.5, 118.8, 118.9, 127.9, 128.9, 129.1, 129.8, 129.9, 130.0, 132.6, 132.7, 132.8, 139.5, 140.0, 141.3, 148.3, 148.4, 148.6, 153.0; FABMS m/z 528 (M<sup>+</sup>+H). Anal. Calcd for C<sub>29</sub>H<sub>33</sub>N<sub>5</sub>O<sub>3</sub>Si: C, 66.01; H, 6.30; N, 13.27. Found: C, 65.99; H, 6.18; N, 13.27.

## 4.18. $(\pm)$ -9-[c-4-(tert-Butyldiphenylsilyloxy)methylt-4,t-5-bis(hydroxymethyl)-cyclopent-2-en-r-1-yl]-9Hadenine (**31**)

To a MeOH (40 mL) solution of **30** (200 mg, 0.38 mmol) was added NaBH<sub>4</sub> (43 mg, 1.14 mmol) at 0 °C. After stirring for 1 h at room temperature, the reaction mixture was partitioned between 0.5 M HCl and CHCl<sub>3</sub>. Silica gel column chromatography (CHCl<sub>3</sub>/MeOH=20:1) of the organic layer gave **31** (173 mg, 86%) as an oil.

<sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.00 (9H, s, SiBu-*tert*), 2.56–2.62 (1H, m, H-5'), 3.57–3.83 (6H, m, CH<sub>2</sub>OSi and CH<sub>2</sub>OH), 4.58 (1H, t, *J*=4.8 Hz, OH), 4.70 (1H, t, *J*=5.0 Hz, OH), 5.42–5.45 (1H, m, H-1'), 5.92 (1H, dd, *J*=5.7 and 1.5 Hz, H-2' or H-3'), 6.00 (1H, dd, *J*=5.7 and 2.1 Hz, H-2' or H-3'), 7.20 (2H, br, NH<sub>2</sub>), 7.39–7.48 (6H, m, Ph), 7.61–7.64 (4H, m, Ph), 7.94 (1H, s, H-8), 8.09 (1H, s, H-2); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  18.7, 26.4, 52.5, 57.6, 59.0, 61.6, 62.1, 66.8, 118.6, 127.6, 127.7, 129.6, 130.2, 130.7, 132.7, 134.9, 138.4, 138.8, 149.3, 152.1, 155.7; FABMS *m*/*z* 530 (M<sup>+</sup>+H). Anal.

Calcd for  $C_{29}H_{35}N_5O_3Si \cdot 1/5H_2O$ : C, 65.31; H, 6.69; N, 13.13. Found: C, 65.02; H, 6.70; N, 12.90.

## 4.19. $(\pm)$ -9-[c-4-(tert-Butyldiphenylsilyloxy)methyl-t-5hydroxymethyl-t-4-vinylcyclopent-2-en-r-1-yl]-9Hadenine (32)

To a suspension of methyltriphenylphosphonium bromide (1.04 g, 2.91 mmol) in THF (30 mL) was added dropwise BuLi (1.67 M in hexane, 1.57 mL, 2.62 mmol) at -78 °C. The resulting suspension was allowed to warm to 0 °C and stirred for 1 h. To this, a THF (4 mL) solution of **30** (154 mg, 0.29 mmol) was added at -78 °C. The reaction mixture was stirred at room temperature for 12 h, and then partitioned between saturated aqueous NH<sub>4</sub>Cl and Et<sub>2</sub>O. Silica gel column chromatography (CHCl<sub>3</sub>/MeOH=80:1) of the organic layer gave **32** (146 mg, 96%) as a foam.

<sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.00 (9H, s, SiBu-*tert*), 2.68–2.73 (1H, m H-5'), 3.50–3.54 (1H, m, CH<sub>2</sub>OH), 3.60–3.65 (1H, m, CH<sub>2</sub>OH), 3.82 (1H, d, *J*=9.7 Hz, CH<sub>2</sub>OSi), 3.84 (1H, d, *J*=9.7 Hz, CH<sub>2</sub>OSi), 4.56 (1H, t, *J*=4.6 Hz, OH), 5.01 (1H, dd, *J*=17.8 and 1.7 Hz, CH=CH<sub>2</sub>), 5.19 (1H, dd, *J*=10.6 and 1.7 Hz, CH=CH<sub>2</sub>), 5.23 (1H, d, *J*=8.0 Hz, H-1'), 5.98 (1H, dd, *J*=17.8 and 10.6 Hz, CH=CH<sub>2</sub>), 6.02–6.04 (2H, m, H-2' and H-3'), 7.24 (2H, br, NH<sub>2</sub>), 7.40–7.48 (6H, m, Ph), 7.61–7.64 (4H, m, Ph), 7.97 (1H, s, H-8), 8.08 (1H, s, H-2); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  19.0, 26.6, 53.4, 58.8, 60.5, 61.7, 68.8, 115.9, 118.9, 127.9, 129.8, 129.9, 130.6, 132.9, 135.2, 138.4, 138.6, 138.8, 149.4, 152.3, 156.0; FABMS *m*/*z* 526 (M<sup>+</sup>+H). Anal. Calcd for C<sub>30</sub>H<sub>35</sub>N<sub>5</sub>O<sub>2</sub>Si·1/10H<sub>2</sub>O: C, 68.31; H, 6.73; N, 13.28. Found: C, 68.04; H, 6.72; N, 13.11.

## 4.20. $(\pm)$ -c-4-(Adenine-9-yl)-1-(tert-butyldiphenylsilyloxy)methyl-7-oxabicyclo-[3.3.0]oct-2-en-8-ylacetic acid methyl ester (33)

A mixture of **30** (38 mg, 0.07 mmol) and  $Ph_3P$ = CHCO<sub>2</sub>Me (60 mg, 0.18 mmol) in xylene (8 mL) was refluxed for 24 h. The reaction mixture was partitioned between H<sub>2</sub>O and CHCl<sub>3</sub>. Silica gel column chromatography (CHCl<sub>3</sub>/ MeOH=50:1) of the organic layer gave **33** (30 mg, 72%, a single isomer, stereochemistry not known) as a foam.

<sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.06 (9H, s, SiBu-*tert*), 2.45–2.47 (2H, m, H-5' and CHCH<sub>2</sub>CO<sub>2</sub>Me), 2.68 (1H, dd, *J*=15.2 and 4.0 Hz, CHCH<sub>2</sub>CO<sub>2</sub>Me), 3.71 (3H, s, OMe), 3.78–3.84 (3H, m, CH<sub>2</sub>OSi and CH<sub>2</sub>O–), 4.21 (1H, dd, *J*=9.7 and 4.0 Hz, CHCH<sub>2</sub>CO<sub>2</sub>Me), 4.29 (1H, d, *J*=9.7 Hz, CH<sub>2</sub>O–), 5.49–5.51 (1H, m, H-1'), 5.76 (2H, br, NH<sub>2</sub>), 5.95 (1H, dd, *J*=5.7 and 2.2 Hz, CH=CH), 6.03 (1H, dd, *J*=5.7 and 2.2 Hz, CH=CH), 7.34–7.47 (6H, m, Ph), 7.59–7.63 (5H, m, Ph and H-8), 8.30 (1H, s, H-2); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  19.2, 26.9, 37.1, 51.9, 55.6, 66.6, 67.1, 67.5, 72.8, 80.2, 120.0, 127.8, 130.0, 130.8, 132.6, 135.5, 135.6, 137.3, 138.1, 149.8, 153.0, 153.7, 155.3, 171.5; FABMS *m*/*z* 584 (M<sup>+</sup>+H). Anal. Calcd for C<sub>32</sub>H<sub>37</sub>N<sub>5</sub>O<sub>4</sub>Si: C, 65.84; H, 6.39; N, 12.00. Found: C, 65.85; H, 6.39; N, 12.12.

4.21. ( $\pm$ )-9-[t-5-Acetoxymethyl-c-4-(tert-butyldiphenylsilyloxy)methyl-t-4-cyanocyclopent-2-en-r-1-yl]-9H-adenine (**34**)

A mixture of **30** (197 mg, 0.37 mmol) and NH<sub>2</sub>OH·HCl (520 mg, 7.48 mmol) in pyridine (5 mL) was stirred at room temperature for 3 h. The reaction mixture was partitioned between saturated aqueous NaHCO<sub>3</sub> and CHCl<sub>3</sub>. The organic layer was evaporated to leave the crude oxime. The crude oxime was dissolved in MeCN (12 mL) and reacted with Ac<sub>2</sub>O (106  $\mu$ L, 1.12 mmol) in the presence of *i*-Pr<sub>2</sub>NEt (195  $\mu$ L, 1.12 mmol) and DMAP (137 mg, 1.12 mmol) for 1 h. The reaction mixture was partitioned between 0.5 M HCl and CHCl<sub>3</sub>. After evaporation of the organic layer, the residue was dissolved in AcOH (6 mL) containing NaOAc (27 mg, 0.33 mmol) and the whole mixture was heated at 100 °C for 3 h. The reaction mixture was evaporated, and then partitioned between saturated aqueous NaHCO<sub>3</sub> and CHCl<sub>3</sub>. Silica gel column chromatography (CHCl<sub>3</sub>/MeOH=60:1) of the organic layer gave 34 (178 mg, 84% from 30) as a foam. IR (neat) 2236 cm<sup>-1</sup> (C $\equiv$ N); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.10 (9H, s, SiButert), 1.83 (3H, s, Ac), 2.94 (1H, dd, J=14.3 and 7.4 Hz, H-5'), 3.93 (1H, d, J=9.7 Hz, CH<sub>2</sub>OSi), 4.01 (1H, d, J=9.7 Hz, CH2OSi), 4.49-4.58 (2H, m, CH2OAc), 5.67-5.69 (1H, m, H-1'), 6.02 (2H, br, NH<sub>2</sub>), 6.10 (1H, dd, J=5.7 and 1.7 Hz, CH=CH), 6.14 (1H, dd, J=5.7 and 2.3 Hz, CH=CH), 7.27-7.49 (6H, m, Ph), 7.65-7.70 (5H, m, Ph and H-8), 8.26 (1H, s, H-2); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 19.3, 20.4, 26.7, 49.1, 53.3, 62.3, 63.4, 67.2, 117.8, 119.8, 127.9, 130.2, 132.0, 132.1, 133.2, 133.5, 135.5, 135.6, 138.4, 149.8, 153.1, 155.6, 170.2; FABMS m/z 567 (M<sup>+</sup>+H). Anal. Calcd for C<sub>31</sub>H<sub>34</sub>N<sub>6</sub>O<sub>3</sub>Si· 3/10H2O: C, 65.08; H, 6.10; N, 14.69. Found: C, 64.86; H, 5.93; N, 14.54.

## 4.22. (±)-9-[c-4-(tert-Butyldiphenylsilyloxy)methyl-t-4ethynyl-t-5-hydroxymethylcyclopent-2-en-r-1-yl]-9Hadenine (**35**)

To a LDA solution in THF (30 mL), prepared from diisopropylamine (626 µL, 4.47 mmol) and BuLi (1.66 M in hexane, 2.53 mL, 4.2 mmol) at 0 °C, TMSCHN<sub>2</sub> (0.6 M in hexane, 7.45 mL, 4.47 mmol) was added at -78 °C. The mixture was stirred at -78 °C for 0.5 h. Compound **30** (295 mg, 0.56 mmol) in THF (30 mL) was added to the mixture at -78 °C. The whole mixture was stirred for 0.5 h at -78 °C and then for 3 h at room temperature. The reaction was quenched by adding saturated aqueous NH<sub>4</sub>Cl. Extraction with CHCl<sub>3</sub> followed by preparative TLC (CHCl<sub>3</sub>/MeOH= 20:1) gave **35** (92 mg, 31%) as a foam. IR (neat)  $2220 \text{ cm}^{-1}$  $(C \equiv C)$ ; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.05 (9H, s, SiBu-tert), 2.38 (1H, s, C≡CH), 2.57-2.62 (1H, m, H-5'), 3.76 (1H, d, J=10.0 Hz, CH<sub>2</sub>OSi), 3.79 (1H, d, J=10.0 Hz, CH<sub>2</sub>OSi), 3.80 (1H, br, OH), 3.97-4.07 (2H, m, CH<sub>2</sub>OH), 5.62-5.64 (1H, m, H-1'), 5.68 (2H, br, NH<sub>2</sub>), 5.96 (1H, dd, J=5.6 and 2.0 Hz, H-2'), 6.04 (1H, dd, J=5.6 and 2.0 Hz, H-3'), 7.34-7.45 (6H, m, Ph), 7.60-7.65 (4H, m, Ph), 7.75 (1H, s, H-8), 8.35 (1H, s, H-2); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 19.3, 26.9, 52.6,

53.8, 62.6, 62.9, 68.9, 74.1, 81.8, 119.9, 127.8, 129.2, 129.9, 132.7, 135.5, 135.6, 138.3, 138.9, 149.7, 152.9, 155.6; FABMS *m*/*z* 524 (M<sup>+</sup>+H). Anal. Calcd for  $C_{30}H_{33}N_5O_2Si$ : C, 68.80; H, 6.35; N, 13.37. Found: C, 68.54; H, 6.40; N, 13.35.

## 4.23. $(\pm)$ -9-[c-4,t-4,t-5-Tris(hydroxymethyl)cyclopent-2-en-r-1-yl]-9H-adenine (**36**)

To a THF (8 mL) solution of **31** (154 mg, 0.291 mmol) was added Bu<sub>4</sub>NF (1 M solution in THF, 320 µL, 0.32 mmol). After being stirred for 0.5 h, the reaction mixture containing the free nucleoside was reacted with Ac<sub>2</sub>O (110 µL, 1.16 mmol) in the presence of *i*-Pr<sub>2</sub>NEt (210 µL, 0.32 mmol) and DMAP (143 mg, 1.16 mmol) for 1 h. The reaction mixture was partitioned between saturated aqueous NaHCO<sub>3</sub> and CHCl<sub>3</sub>. Silica gel column chromatography (CH<sub>2</sub>Cl<sub>2</sub>/MeOH=100:1) of the organic layer gave the triacetate (99 mg). The triacetate was treated with NH<sub>3</sub>/MeOH (5 mL) at 5 °C for 24 h. After evaporation, the residue was purified by silica gel column chromatography (CHCl<sub>3</sub>/MeOH=40:1). This gave **36** (48 mg, 74%) as a foam.

<sup>1</sup>H NMR (CD<sub>3</sub>OD)  $\delta$  2.54–2.60 (1H, m, H-5'), 3.61–3.96 (6H, m, CH<sub>2</sub>OH), 5.58–5.61 (1H, m H-1'), 5.95 (1H, dd, J=5.6 and 1.7 Hz, H-2' or H-3'), 6.00 (1H, dd, J=5.6 and 2.1 Hz, H-2' or H-3'), 8.17 and 8.18 (2H, each as s, H-8 and H-2); <sup>13</sup>C NMR (DMSO- $d_6$ )  $\delta$  53.8, 57.9, 59.1, 61.9, 62.3, 65.3, 118.9, 130.2, 139.1, 139.2, 149.5, 152.2, 155.9; FABMS m/z 292 (M<sup>+</sup>+H). Anal. Calcd for C<sub>13</sub>H<sub>17</sub>N<sub>5</sub>O<sub>3</sub>·1/5H<sub>2</sub>O: C, 52.95; H, 5.95; N, 23.75. Found: C, 53.18; H, 6.03; N, 23.51.

## 4.24. $(\pm)$ -9-[c-4,t-5-Bis(hydroxymethyl)-t-4vinylcyclopent-2-en-r-1-yl]-9H-adenine (**37**)

Compound **37** (42 mg, 69%) was obtained as a foam from **32** (113 mg, 0.214 mmol) by the procedure described for the preparation of **36**. <sup>1</sup>H NMR (CD<sub>3</sub>OD)  $\delta$  2.65–2.70 (1H, m, H-5'), 3.63–3.81 (4H, m, CH<sub>2</sub>OH), 5.12 (1H, dd, *J*=17.6 and, 1.5 Hz, CH=CH<sub>2</sub>), 5.25 (1H, dd, *J*=10.7 and 1.5 Hz, CH=CH<sub>2</sub>), 5.39 (1H, d, *J*=7.6 Hz, H-1'), 5.96–6.03 (3H, m, H-2', H-3' and CH=CH<sub>2</sub>), 8.18 (1H, s, H-8), 8.20 (1H, s, H-2); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>)  $\delta$  55.7, 60.5, 62.3, 64.1, 68.7, 117.0, 120.1, 131.3, 138.8, 140.7, 141.2, 150.6, 153.6, 157.3; FABMS *m*/*z* 288 (M<sup>+</sup>+H). Anal. Calcd for C<sub>14</sub>H<sub>17</sub>N<sub>5</sub>O<sub>2</sub>·1/10H<sub>2</sub>O: C, 58.16; H, 6.00; N, 24.22. Found: C, 58.08; H, 5.94; N, 24.48.

## 4.25. $(\pm)$ -9-[t-4-Ethynyl-c-4,t-5-bis(hydroxymethyl)cyclopent-2-en-r-1-yl]-9H-adenine (38)

Compound **38** (44 mg, 70%, solid) was obtained from **35** (116 mg, 0.22 mmol) by the procedure described for the preparation of **36**. Mp 113–115 °C; IR (KBr) 2100 cm<sup>-1</sup> (C $\equiv$ C); <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>)  $\delta$  2.60 (1H, q, *J*=7.5 Hz, H-5'), 3.16 (1H, s, ethynyl), 3.56–3.59 (1H, m, C*H*<sub>2</sub>OH), 3.62–3.67 (2H, m, *CH*<sub>2</sub>OH), 3.82–3.87 (1H, m, *CH*<sub>2</sub>OH), 4.61–4.63 (1H, m, OH), 5.21–5.24 (1H, m, OH), 5.37 (1H, dt, *J*=7.5 and 2.3 Hz, H-1'), 5.88 (1H, dd, *J*=5.2 and 2.3 Hz, H-2' or

H-3'), 5.97 (1H, dd, J=5.2 and 2.3 Hz, H-2' or H-3'), 7.19 (2H, br, NH<sub>2</sub>), 8.05 (1H, s, H-8), 8.11 (1H, s, H-2); <sup>13</sup>C NMR (DMSO- $d_6$ )  $\delta$  52.2, 52.4, 61.4, 62.3, 66.8, 75.7, 82.9, 118.8, 130.4, 137.3, 139.1, 149.2, 152.2, 155.9; FABMS m/z 286 (M<sup>+</sup>+H). Anal. Calcd for C<sub>14</sub>H<sub>15</sub>N<sub>5</sub>O<sub>2</sub>·3/4H<sub>2</sub>O: C, 56.11; H, 5.58; N, 23.37. Found: C, 56.30; H, 5.35; N, 23.29.

## 4.26. (±)-c-4-(Adenine-9-yl)-1-hydroxymethyl-7oxabicyclo-[3.3.0]oct-2-en-8-ylideneamine (**39**)

To a THF (8 mL) solution of 34 (135 mg, 0.238 mmol) was added Bu<sub>4</sub>NF (1 M solution in THF, 262 µL, 0.262 mmol). After being stirred for 1 h, the reaction mixture containing the free nucleoside was reacted with  $Ac_2O$  (34 µL, 0.36 mmol) in the presence of *i*-Pr<sub>2</sub>NEt (104 µL, 0.59 mmol) and DMAP (44 mg, 0.36 mmol) for 1 h. The reaction mixture was partitioned between saturated aqueous NaHCO<sub>3</sub> and CHCl<sub>3</sub>. Silica gel column chromatography (CH<sub>2</sub>Cl<sub>2</sub>/MeOH=40:1) of the organic layer gave the diacetate (87 mg). The acetate was treated with NH<sub>3</sub>/MeOH (5 mL) at 5 °C for 18 h. During evaporation of the solvent, precipitation occurred. The precipitate was washed with hot benzene (50 mL) to give an analytically pure 39 (38 mg, 79%, solid, mixture of two isomers ca. 6:4). Mp 237 °C; <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>) δ 3.06-3.11 (1H, m, H-5'), 3.66-3.76 (2H, m, CH<sub>2</sub>OH), 4.24-4.29 (1H, m, CH<sub>2</sub>O), 4.41-4.47 (1H, m, CH<sub>2</sub>O), 5.01-5.02 (0.6H, m, OH), 5.14-5.16 (0.4H, m, OH), 5.49-5.51 (1H, m, H-1'), 5.99-6.02 (1.6H, m, CH=CH), 6.09-6.11 (0.4H, m, CH=CH), 7.25 (2H, br, NH<sub>2</sub>), 7.63 (0.6H, s, NH), 7.93 (0.4H, s, NH), 8.01 (0.4H, s, H-8), 8.03 (0.6H, s, H-8), 8.15 (1H, s, H-2); <sup>13</sup>C NMR (DMSO- $d_6$ )  $\delta$  50.6, 50.7, 63.9, 64.4, 64.5, 65.5, 66.1, 71.5, 72.1, 119.0, 119.1, 129.2, 129.4, 136.8, 137.9, 138.7, 138.8, 149.1, 152.4, 156.0, 169.9, 174.6; FABMS m/z 287  $(M^++H)$ . Anal. Calcd for  $C_{13}H_{14}N_6O_2 \cdot 1/10H_2O$ : C, 54.20; H, 4.97; N, 29.17. Found: C, 54.13; H, 4.89; N, 29.11.

#### 4.27. Anti-HIV-1 assay

MT-4 cells  $(1 \times 10^5 \text{ cells/mL})$  were infected with HIV-1 (HTLV-III<sub>B</sub> strain) at a multiplicity of infection (MOI) of 0.02 and were cultured in the presence of various concentrations of the test compounds. After a 4-day incubation at 37 °C, the number of viable cells was monitored by the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide method. The cytotoxicity of the compounds was evaluated in parallel with their antiviral activity, based on the viability of mock-infected cells.

#### 4.28. Anti-HCV assay

OR6 cells, a cell line cloned from ORN/C-5B/KE cells22) that supports genome-length HCV RNA (strain O of genotype 1b) encoding *Renilla* luciferase reporter gene, were cultured in Dulbecco's modified Eagle's medium supplemented with 10% fetal bovine serum in the presence of G418 (300 mg/mL: Geneticin; Invitrogen, Carlsbad, CA). To monitor the anti-HCV effects of the compounds, OR6 cells were plated onto

24-well plates in triplicate  $(1.5 \times 10^4 \text{ cells per well})$  and cultured for 24 h. Then, the cells were treated with the compounds (10  $\mu$ M each) for 72 h. After treatment, the cells were harvested with *Renilla* luciferase lysis reagent (Promega, Madison, WI) and subjected to the *Renilla* luciferase assay according to the manufacturer's protocol.

### Acknowledgements

Financial support from the Japan Society for the Promotion of Science (KAKENHI, Grant no. 18790090 to H.K.) is gratefully acknowledged. The authors are also grateful to Ms. K. Shiobara and Ms. Y. Odanaka (Center for Instrumental Analysis, Showa University) for technical assistance with NMR, MS, and elemental analysis. T.H. is a Research Resident of the Japanese Foundation for AIDS Prevention.

#### **References and notes**

- (a) Vince, R.; Hua, M. J. Med. Chem. 1990, 33, 17; (b) Daluge, S.M. Therapeutic Nucleosides. U.S. Patent 5,034,394, 1991.
- 2. Huff, J. R. Bioorg. Med. Chem. 1999, 7, 2667.
- (a) Katagiri, N.; Nomura, M.; Sato, H.; Kaneko, C.; Yusa, K.; Tsuruo, T. J. Med. Chem. 1992, 35, 1882; (b) Katagiri, N.; Shiraishi, T.; Sato, H.; Toyota, A.; Kaneko, C.; Yusa, K.; Ohhara, T.; Tsuruo, T. Biochem. Biophys. Res. Commun. 1992, 184, 154; (c) Katagiri, N.; Shiraishi, T.; Toyota, A.; Sato, H.; Kaneko, C.; Aikawa, T. Chem. Pharm. Bull. 1993, 41, 1027; (d) Tanaka, M.; Norimine, Y.; Fujita, T.; Suemune, H. J. Org. Chem. 1996, 61, 6952; (e) Banerjee, S.; Ghosh, S.; Shinha, S. J. Org. Chem. 2005, 70, 4199.
- (a) Norbeck, D. W.; Kern, E.; Hayashi, S.; Rosenbrook, W.; Sham, H.; Herrin, T.; Plattner, J. J.; Erickson, J.; Clement, J.; Swanson, R.; Shipkowitz, N.; Hardy, D.; Marsh, K.; Arnett, G.; Shanmon, W.; Broder, S.; Mitsuya, H. J. Med. Chem. 1990, 33, 1281; (b) Bisaochi, B. S.; Braitman, A.; Cianci, C. W.; Clark, J. M.; Field, A. K.; Hagen, M. E.; Hochstein, D. R.; Malley, M. F.; Mitt, T.; Slusarchyk, W. A.; Sundeen, J. E.; Terry, B. J.; Tuomari, A. V.; Weaver, E. R.; Yung, M. G.; Zahler, R. J. Med. Chem. 1991, 34, 1415.
- Shimada, N.; Hasegawa, S.; Harada, T.; Tomisawa, T.; Fujii, A.; Takita, T. J. Antibiot. 1986, 34, 1623.
- (a) Prisbe, E. J.; Magg, H.; Verheyden, J. P. H.; Rydzewski, R. M. Nucleosides and Nucleotides as Antitumor and Antiviral Agents; Chu, C. K., Baker, D. C., Eds.; Plenum: New York, NY, 1993; p 101; (b) Kodama, E.; Kohgo, S.; Kitano, K.; Machida, H.; Gatanaga, H.; Shigeta, S.; Matsuoka, M.; Ohrui, H.; Mitsuya, H. Antimicrob. Agents Chemother. 2001, 45, 1539; (c) Hayakawa, H.; Kohgo, S.; Kitano, K.; Ashida, N.; Kodama, E.; Mitsuya, H.; Ohrui, H. Antiviral Chem. Chemother. 2004, 15, 169.

- (a) Haraguchi, K.; Takeda, S.; Tanaka, H.; Nitanda, T.; Baba, M.; Dutschman, G. E.; Cheng, Y.-C. *Bioorg. Med. Chem. Lett.* 2003, 13, 3775; (b) Dutschman, G. E.; Grill, S. P.; Gullen, E. A.; Haraguchi, K.; Takeda, S.; Tanaka, H.; Baba, M.; Cheng, Y.-C. *Antimicrob. Agents Chemother.* 2004, 48, 1640; (c) Nitanda, T.; Wang, X.; Kumamoto, H.; Haraguchi, K.; Tanaka, H.; Cheng, Y.-C.; Baba, M. *Antimicrob. Agents Chemother.* 2005, 49, 3355; (d) Tanaka, H.; Haraguchi, K.; Kumamoto, H.; Baba, M.; Cheng, Y.-C. *Antiviral Chem. Chemother.* 2005, 16, 217.
- Mansuri, M. M.; Hitchcock, M. J. M.; Broker, R. A.; Bregman, C. L.; Ghazzouli, L.; Desiderio, J. V.; Starrett, J. E.; Sterzyski, R. Z.; Martin, J. C. Antimicrob. Agents Chemother. 1990, 34, 637.
- For examples of radical-mediated 5-exo-trig cyclization of phenylselenomethyl esters: (a) Beckwith, A. L. J.; Pigou, P. E. J. Chem. Soc., Chem. Commun. 1986, 85; (b) Singh, R.; Singh, G. C.; Ghosh, S. K. Tetrahedron Lett. 2005, 46, 4719.
- Kumamoto, H.; Haraguchi, K.; Tanaka, H.; Nitanda, T.; Baba, M.; Dutschman, G. E.; Cheng, Y.-C.; Kato, K. Nucleosides Nucleotides Nucleic Acids 2005, 24, 73.
- 11. Beckwith, A. L. J.; Pigou, P. E. Aust. J. Chem. 1986, 39, 77.
- 12. A brief calculation was carried out in which A and B are model substrates. Natural bond orbital (NBO) theory was used for analyzing the energy level of molecular orbitals. Our calculations by B3LYP/6-31G\* showed that the π\* of B (0.05644 eV) was located at an energy level slightly lower than that of A (0.06188 eV). This result is suggestive of the lower π\* level of 15 compared with that of 14, and may also contribute for a higher yield production of 16 from 14. (a) Giese, B. Angew. Chem., Int. Ed. Engl. 1983, 22, 753; (b) Fleming, I. Frontier Orbitals and Organic Chemical Reactions; Wiley: London, 1976.



- (a) Newcomb, M. Tetrahedron 1993, 49, 1151; (b) Chatgilialoglu, C.; Alberti, A.; Ballestri, M.; Macciantelli, D. Tetrahedron Lett. 1996, 37, 6391.
- Andrau, L.; Lebreton, J.; Viazzo, P.; Alphand, V.; Furstoss, R. *Tetrahedron* Lett. 1997, 38, 825.
- 15. Ghosh, S.; Shinha, S.; Drew, M. G. B. Org. Lett. 2006, 8, 3781.
- 16. Trost, B. M. J. Org. Chem. 2004, 69, 5813.
- (a) Day, S.; Garner, P. J. Org. Chem. 2000, 65, 7697; (b) Yin, X.; Li, W.; Schneller, S. W. Tetrahedron Lett. 2006, 47, 9187.
- 18. Matsumoto, M.; Kuroda, K. Tetrahedron Lett. 1980, 21, 4021.
- 19. Corey, E. J.; Ruden, R. A. Tetrahedron Lett. 1973, 17, 1495.
- (a) Ohira, S. Synth. Commun. 1989, 19, 561; (b) Rauhala, V.; Nevalainen, M.; Koskinen, A. M. P. Tetrahedron 2004, 60, 9199.
- 21. Colvin, E. W.; Hamill, B. J. J. Chem. Soc., Chem. Commun. 1973, 151.
- Ikeda, M.; Abe, K.; Dansako, H.; Nakamura, T.; Naka, K.; Kato, N. Biochem. Biophys. Res. Commun. 2005, 329, 1350.
- Naka, K.; Ikeda, M.; Abe, K.; Dansako, H.; Kato, N. Biochem. Biophys. Res. Commun. 2005, 330, 871.